

• 综述 •

miRNA 相关抗结核免疫调控及其潜在治疗靶点的研究进展

周妍阳, 胥萍*

苏州大学苏州医学院, 江苏 苏州 215131

[摘要] 微小RNA (microRNA, miRNA) 是调控转录后基因表达的一类非编码RNA, 具有调控多种细胞生物学过程的功能。越来越多的研究表明, 结核分枝杆菌 (*Mycobacterium tuberculosis*, Mtb) 感染后宿主细胞内多种 miRNA 发生差异表达, 进而通过影响下游通路来调控宿主抗结核 (tuberculosis, TB) 免疫。文章就 Mtb 感染后 miRNA 含量改变对下游因子的调控以及对自噬、凋亡和炎症反应的影响进行综述, 认为 miRNA 是一种有潜力的 TB 治疗靶点, 旨在为 miRNA 在 TB 中的深入研究和临床应用提供参考依据。

[关键词] miRNA; 结核病; 自噬; 凋亡; 治疗靶点

[中图分类号] R52

[文献标志码] A

[文章编号] 1007-4368(2024)12-1745-11

doi: 10.7655/NYDXBNSN240332

Research progress of miRNA-related anti-tuberculosis immune regulation and its potential therapeutic targets

ZHOU Yanyang, XU Ping*

Suzhou Medical College, Soochow University, Suzhou 215131, China

[Abstract] MicroRNAs (miRNA) are a class of non-coding RNA molecules that regulate gene expression transcriptionally, playing crucial roles in various cellular processes. Increasing investigation indicate that the *Mycobacterium tuberculosis* (Mtb) infection alters the expression of numerous miRNAs in host cells, thereby influencing downstream pathways involved in immune responses against tuberculosis (TB). This review summarizes how changes in miRNA levels post Mtb infection regulate autophagy, apoptosis, and inflammatory responses. It highlights that miRNAs may serve as potential therapeutic targets for TB, providing insights for further research and clinical applications of miRNA in TB.

[Key words] miRNA; tuberculosis; autophagy; apoptosis; therapeutic target

[J Nanjing Med Univ, 2024, 44(12): 1745-1754, 1762]

结核病 (tuberculosis, TB) 是由结核分枝杆菌 (*Mycobacterium tuberculosis*, Mtb) 引起的一种经由呼吸道传播的传染性疾病。根据世界卫生组织发布的《2023年全球结核病报告》, TB 是仅次于新型冠状病毒肺炎的全球第二大单一传染病死亡病因^[1]。Mtb 主要感染肺部, 导致典型的肺结核综合征^[2]。宿主的免疫功能参与细胞的自噬、凋亡和吞噬溶酶体成熟等生物学过程, 对控制 Mtb 感染至关重要。越来

[基金项目] 江苏省科技计划专项 (BE2023718); 江苏省共建放射医学与辐射防护国家重点实验室开放研究课题 (GZK12023015)

*通信作者 (Corresponding author), E-mail: 573311485@qq.com

越多的证据表明, 微小RNA (microRNA, miRNA) 可调控上述大多数生物学过程。miRNA 是长度为 18~25 个核苷酸的小分子 RNA, 不参与蛋白质的编码, 而在细胞增殖分化、信号通路激活、免疫功能调节等生物学过程中充当基因表达的关键控制者^[3]。miRNA 通过 Argonaute (AGO) 蛋白形成 RNA 诱导的基因沉默复合物 (RNA-induced silencing complex, RISC) 沉默靶 mRNA 的表达^[4], 对靶基因的表达产生负调控作用。

近年来, 越来越多的研究发现 TB 患者和健康人存在 miRNA 之间的表达差异, 进而探究出差异表达的 miRNA 通过影响下游通路在 Mtb 感染细胞中导

致细胞分子层面的变化(如自噬受损、凋亡受损、促炎细胞因子产生受阻等)。进一步提出miRNA可作为TB的潜在治疗靶点,并通过体内实验进行验证。文章就miRNA相关抗TB免疫调控及其潜在治疗靶点进行综述,旨在为miRNA在TB中的深入研究和临床应用提供参考依据。

1 Mtb感染后miRNA的差异表达调控抗TB免疫

1.1 miRNA对自噬的调控

自噬是胞质内容物被质膜包裹形成自噬体后转运至溶酶体,由溶酶体降解内容物的过程^[5],已被证明可以增强对胞内病原菌的杀伤^[6]。通过自噬清除胞内病原体的过程被称为异噬,是宿主对抗多种病原体的先天性防御策略^[7]。自噬相关通路在转录后水平受到严格的调控,但在Mtb感染期间,miRNA对自噬的影响在很大程度上是未知的。近几年的研究证实Mtb感染期间miRNA的差异表达对自噬存在调控作用(图1)。

1.1.1 miRNA调控自噬相关基因(autophagy related gene, ATG)

几乎所有的ATG蛋白均参与自噬的生物学过程^[8],miRNA可通过靶向ATG的3'非编码区发挥抑制作用,从而对自噬产生影响。在Mtb感染后,miRNA-142-3p在巨噬细胞内的含量下调,其靶基因ATG4c和ATG16L1上调,从而促进了细胞自噬^[9];感染Mtb的巨噬细胞中miRNA-106a显著下调,从而解除其对Unc-51样自噬激活激酶(Unc-51 like autophagy activating kinase 1, ULK1)和ATG7/ATG16L1的抑制,促进自噬^[10]。相似地,Mtb感染的巨噬细胞中,miRNA-20a显著下调,导致靶基因ATG7和ATG16L1上调以解除miRNA-20a对自噬的抑制^[11]。

目前尚不完全清楚Mtb如何抑制感染细胞的自噬,但miRNA在这一过程中具有重要作用:靶向抑制ATG的miRNA可被Mtb上调,进而抑制自噬,促进Mtb存活。miRNA-144-3p在牛分枝杆菌卡介苗(*Bacillus Calmette-Guérin*, BCG)感染的巨噬细胞中上调,靶向抑制ATG4a,从而抑制自噬,促进BCG的胞内存活^[12]。并且miRNA-144-3p的上调还通过靶向过氧化物酶体增殖物激活受体 α (peroxisome proliferator activated receptor alpha, PPAR α)和三磷酸腺苷结合盒转运体A1(ATP binding cassette subfamily A member 1, ABCA1)促进巨噬细胞的脂质积累,其对宿主脂质代谢的重编程抑制了Mtb感染引起的自噬^[13]。此外,miRNA-155^[14]、miRNA-1958^[15]均被证实

在Mtb感染的细胞中表达上调,并分别通过靶向抑制ATG3和Rheb、ATG5实现对自噬的抑制。

1.1.2 miRNA调控自噬体成熟

自噬体是吞噬泡在ATG蛋白的作用下形成的,其与溶酶体融合,降解胞质内容物,参与胞内病原菌的清除^[8]。Mtb可调控相关miRNA的表达以抑制自噬体的成熟,从而抑制自噬、促进TB进展。miRNA-542-3p在Mtb感染的巨噬细胞中表达上调,通过抑制膜泡分拣蛋白11(vacuolar protein sorting 11, VPS11)抑制自噬体的成熟及其与溶酶体的相互作用,从而抑制细胞自噬^[16]。与上述研究结果类似,miRNA-423-5p在Mtb感染的巨噬细胞中表达升高并抑制其靶基因VPS33A,从而抑制自噬体-溶酶体融合^[17]。Mtb组分Rv1759c可诱导miRNA-25的表达上调,靶向位于溶酶体膜上的细胞内胆固醇转运蛋白C型尼曼-匹克蛋白1(Niemann-Pick C 1, NPC1),导致溶酶体功能受损,从而抑制自噬溶酶体成熟^[18]。此外,毒力蛋白ESAT-6可通过调节miRNA-30a-3p和miRNA-30a-5p的表达水平调控自噬。miRNA-30a-5p的上调会促进感染Mtb的巨噬细胞自噬,而ESAT-6诱导的miRNA-30a-3p通过抑制自噬体和溶酶体的融合从而拮抗miRNA-30a-5p促进自噬的作用^[19]。肿瘤坏死因子样弱凋亡诱导因子(tumor necrosis factor-like weak inducer of apoptosis, TWEAK)可通过激活腺苷酸激活蛋白激酶(adenosine 5'-monophosphate-activated protein kinase, AMPK)诱导自噬体成熟。在Mtb感染的巨噬细胞中,miRNA-889上调靶向抑制TWEAK的表达来抑制自噬^[20]。研究表明,Mtb具有操纵Ca²⁺信号以阻止自噬体成熟和自噬体-溶酶体融合的功能,Mtb可诱导巨噬细胞中的miRNA-27a表达上调,miRNA-27a靶向抑制定位内质网的电压依赖型Ca²⁺通道基因(CACNA203),导致Ca²⁺信号下调,从而抑制自噬体形成^[21]。此外,miRNA-144*在感染的巨噬细胞中上调,靶向抑制DNA损伤调节自噬调节因子2(DNA damage-regulated autophagy modulator protein 2, DRAM2),抑制自噬体的成熟^[22]。与上述相反的是,miRNA-215-5p在TB患者中表达下调,可通过间接靶向SNAP29促进自噬体和溶酶体的融合以促进自噬^[23]。

1.1.3 miRNA调控其他基因影响自噬

差异表达的miRNA也可靶向其他基因,对自噬产生影响。Mmu-miRNA-25-3p在Mtb感染的巨噬细胞中表达上调,其通过抑制双特异性磷酸酶10(dual-specificity protein phosphatase, DUSP10)的表达促进

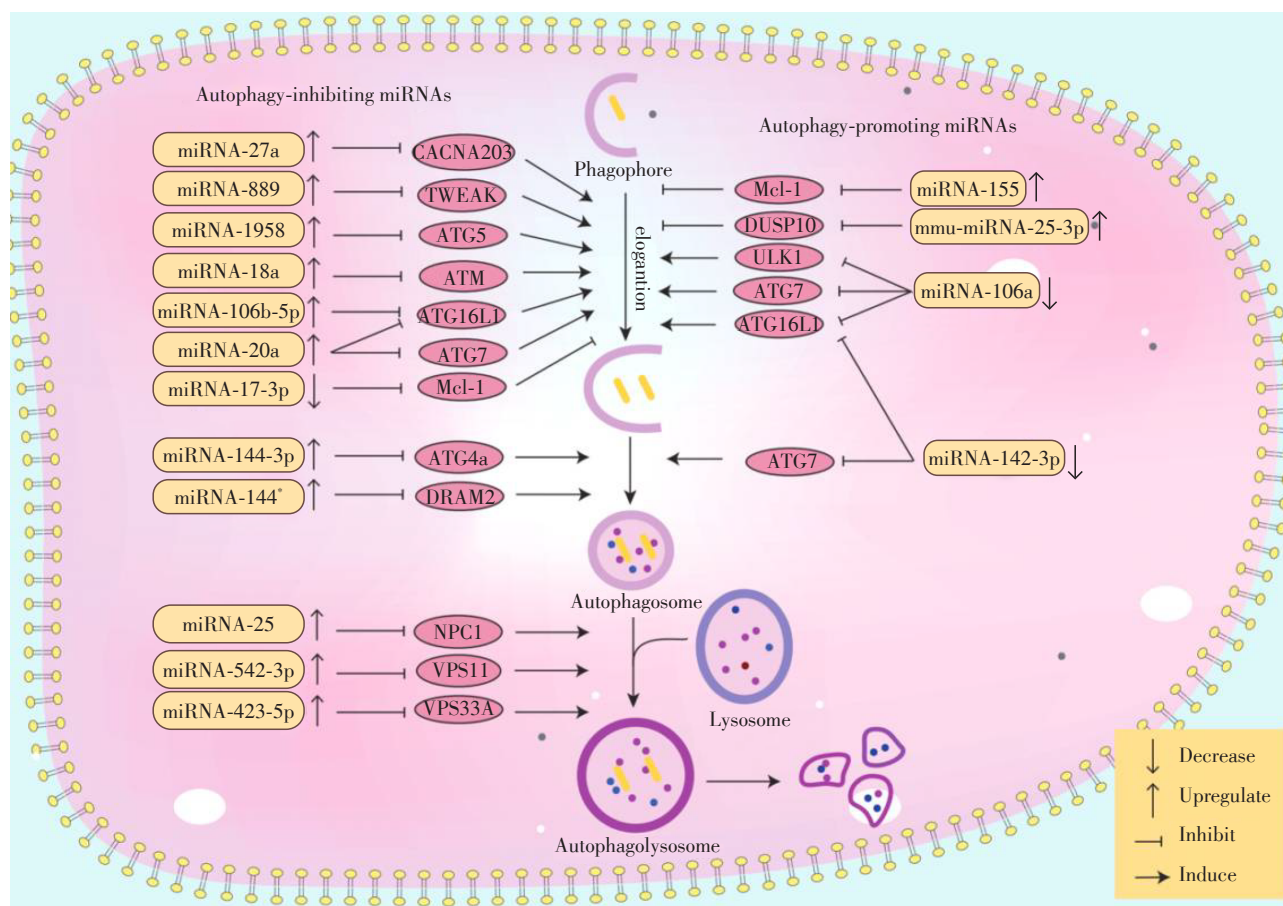


图1 miRNA及其靶基因对自噬的影响

Figure 1 The effect of miRNAs and their target genes on autophagy

ERK1/2 蛋白磷酸化,从而增强 BCG 诱导的巨噬细胞自噬^[24]。miRNA-582-5p 在感染的巨噬细胞中表达上调,靶向抑制端锚聚合酶 2 (tankyrase 2, TNKS2),促进自噬^[25]。毛细血管扩张性共济失调突变(ataxia telangiectasia mutated, ATM)通路可通过激活 LKB1/AMPK/TSC2 信号并抑制负调控因子 mTORC1 以促进自噬。miRNA-18a 在感染 Mtb 的巨噬细胞中上调,并通过下调 ATM 通路抑制自噬过程^[26]。miRNA-17-5p 下调后靶向上调了髓样细胞白血病-1 (myeloid cell leukemia-1, Mcl-1)和信号转导与转录激活因子 3 (signal transducer and activator of transcription 3, STAT3),抑制了 Mtb 诱导的巨噬细胞自噬^[27]。Mcl-1 属于 Bcl-2 家族的抗凋亡成员, Bcl-2 家族成员除了调节凋亡外,也参与自噬的调控^[28]。

总之,自噬对调节 Mtb 感染的免疫反应和巨噬细胞防御至关重要。部分 miRNA 在 Mtb 感染后发生差异表达,影响下游基因调控自噬,进而影响 Mtb 的胞内清除。

1.2 miRNA 对凋亡的调控

细胞凋亡是多种信号共同调控的程序性细胞死亡。小鼠体内实验证实 Fas 诱导的凋亡途径和 Bal-2 调节的凋亡途径分别通过清除感染 Mtb 的巨噬细胞和中性粒细胞在抗结核感染中发挥关键作用^[29]。细胞凋亡可阻止胞内病原体的释放和传播,在宿主防御胞内病原体感染中起到关键作用^[30]。研究表明, Mtb 感染细胞所导致的 miRNA 差异表达,可通过影响凋亡分子的表达进而调控细胞凋亡(图2)。

1.2.1 miR-155 对凋亡调控的双面性

miR-155 在 H37Rv 感染或 EsxA 刺激的巨噬细胞中持续上调^[31]。小鼠在敲除 miRNA-155 后对 Mtb 更易感,且其巨噬细胞凋亡被抑制,说明 miRNA-155 在抗 TB 免疫中的重要作用^[32]。巨噬细胞中的 miRNA-155 在 Mtb 或 BCG 诱导后表达上调,研究表明其上调依赖于 Toll 样受体 (Toll like receptor, TLR)、核因子 κ B (nuclear factor-kappa B, NF- κ B) 和 Jun 氨基末端激酶 (Jun N-terminal kinase, JNK) 信号通路^[31]。miR-155 通过靶向叉形头框蛋白 O3 (forkhead box

O3, FOXO3)抑制单核细胞凋亡^[33]。有趣的是, miRNA-155也可通过靶向SOCS1促进巨噬细胞凋亡^[34]。miRNA-155在Mtb感染时的双面性可能代表了Mtb在不同类型细胞中的抗感染免疫机制,还有待进一步阐明。

1.2.2 其他差异表达miRNA对凋亡的调控

近年来,已有许多研究证明Mtb感染诱导的miRNA差异表达可遏制细胞凋亡。Mtb诱导巨噬细胞中miRNA-325-3p的过表达,从而靶向抑制E3泛素连接酶LNX1(ligand of numb-protein X1),导致中心体相关表达激酶6(NIMA-related expressed kinase 6, NEK6)的异常积累,进而异常激活STAT3信号,抑制细胞凋亡过程^[35]。Mtb感染后,巨噬细胞中miRNA-20b-5p的下调使Mcl-1表达上调以抑制细胞凋亡^[36]。miRNA-223在活动性结核病(active tuberculosis, ATB)患者巨噬细胞中大量表达,通过下调FOXO3抑制巨噬细胞凋亡^[37]。Mtb也可通过调控miRNA诱导宿主细胞死亡途径的转化,促进自身胞内存活。

miRNA-342-3p靶向抑制细胞因子信号抑制因子6(suppressors of cytokine signaling, SOCS6),通过A20介导的K48泛素化和RIPK3降解以及STAT1磷酸化参与Mtb诱导的细胞凋亡和坏死之间的转换。其在感染的细胞中表达下调,促使细胞坏死,Mtb存活^[38]。

Mtb感染后miRNA的差异表达可靶向下游基因促进细胞凋亡。miRNA-27b高表达于感染Mtb的巨噬细胞,通过抑制Bcl-2相关抗凋亡基因2(Bcl-2 associated athanogene 2, Bag2)促进了巨噬细胞p53依赖性细胞凋亡^[39]。BCG感染巨噬细胞后,miRNA-100-5p的下调靶向促进SMARCA e5的上调,进而通过Caspase-3和Bcl-2促进细胞凋亡^[40]。Mtb感染诱导巨噬细胞中的miRNA-20a-5p下调,其靶基因JNK2的表达显著增加,并诱导Bim的表达,随后触发细胞凋亡以促进Mtb清除^[41]。miRNA-125b-5p在感染Mtb的巨噬细胞中上调,可通过靶向DRAM2促进细胞凋亡和抑制炎症反应,从而保护巨噬细胞抵抗Mtb感染^[42]。

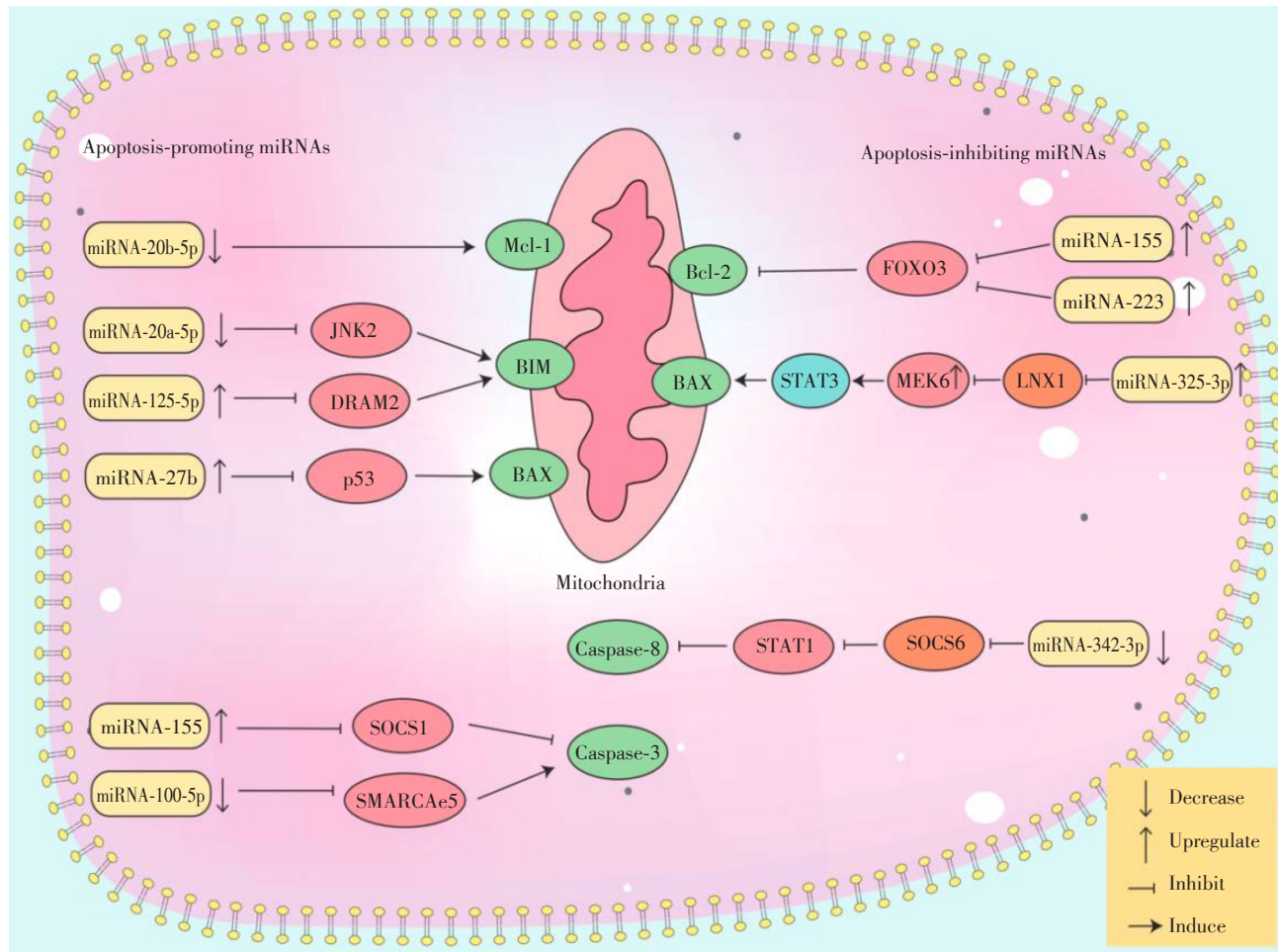


图2 miRNA及其靶基因对凋亡调控因子的影响

Figure 2 The effect of miRNAs and their target genes on apoptosis regulatory factors

综上所述,作为清除胞内菌感染的途径之一,凋亡同自噬一样受到差异表达miRNA的调控,进而影响Mtb的胞内存活。

1.3 miRNA调控促炎细胞因子分泌

炎症是机体对刺激的防御反应,是维持内环境稳定的基本生理过程。当病原体入侵机体后,炎症反应帮助机体清除病原微生物,激发适应性免疫应答^[43],在控制病原体感染中发挥重要作用。

1.3.1 miRNA调控炎症因子分泌

NF- κ B通路直接诱导促炎细胞因子如白细胞介素(interleukin, IL)-1、IL-6、肿瘤坏死因子(tumor necrosis factor, TNF)- α 等的产生,在先天性和适应性免疫应答中发挥重要作用。诱导NF- κ B通路的活化,可显著降低巨噬细胞内的Mtb负荷^[44]。Mtb感染后miRNA表达量的改变可通过影响NF- κ B通路,促进炎细胞因子的释放(图3)。在Mtb感染的巨噬细胞中,miRNA-20a-3p被诱导上调,并下调其靶基因NF- κ B激酶 β 抑制剂(inhibitor of NF- κ B kinase β , IKK β)的表达,从而抑制NF- κ B通路的激活,导致宿主细胞分泌促炎因子以利于Mtb存活^[45]。A20是

NF- κ B信号通路的抑制剂。Mtb可通过抑制miRNA-let-7f的表达,从而上调靶基因A20的表达,抑制促炎因子的分泌^[46]。TNF受体相关因子6(TNF receptor associated factor 6, TRAF6)在NF- κ B通路中作为信号转导子发挥作用,激活IKK β 对促炎细胞因子做出反应。miRNA-146a^[47]和miRNA-125a^[48]在感染细胞中表达上调,靶向TRAF6抑制NF- κ B信号通路,从而抑制相关促炎因子的分泌。Mtb的毒力组分ESAT-6可抑制miRNA-223-3p的合成,导致其靶基因第10号染色体缺失的磷酸酶和张力蛋白同源基因(phosphatase and tensin homolog deleted on chromosome ten, PTEN)上调,抑制PI3K/AKT/NF- κ B通路减少促炎细胞因子的产生^[49]。此外,Mtb感染的巨噬细胞中miRNA-502-3p上调,靶向抑制Rho相关卷曲螺旋含蛋白激酶1(Rho associated coiled-coil containing protein kinase 1, ROCK1)以降低促炎细胞因子的表达。并且过表达miRNA-502-3p显著抑制TLR4/NF- κ B信号通路相关蛋白的表达,可能协同抑制细胞因子的产生^[50]。

与上述相反的是,Mtb感染导致胞内miRNA-340-5p表达显著降低,促进跨膜p24运输蛋白7

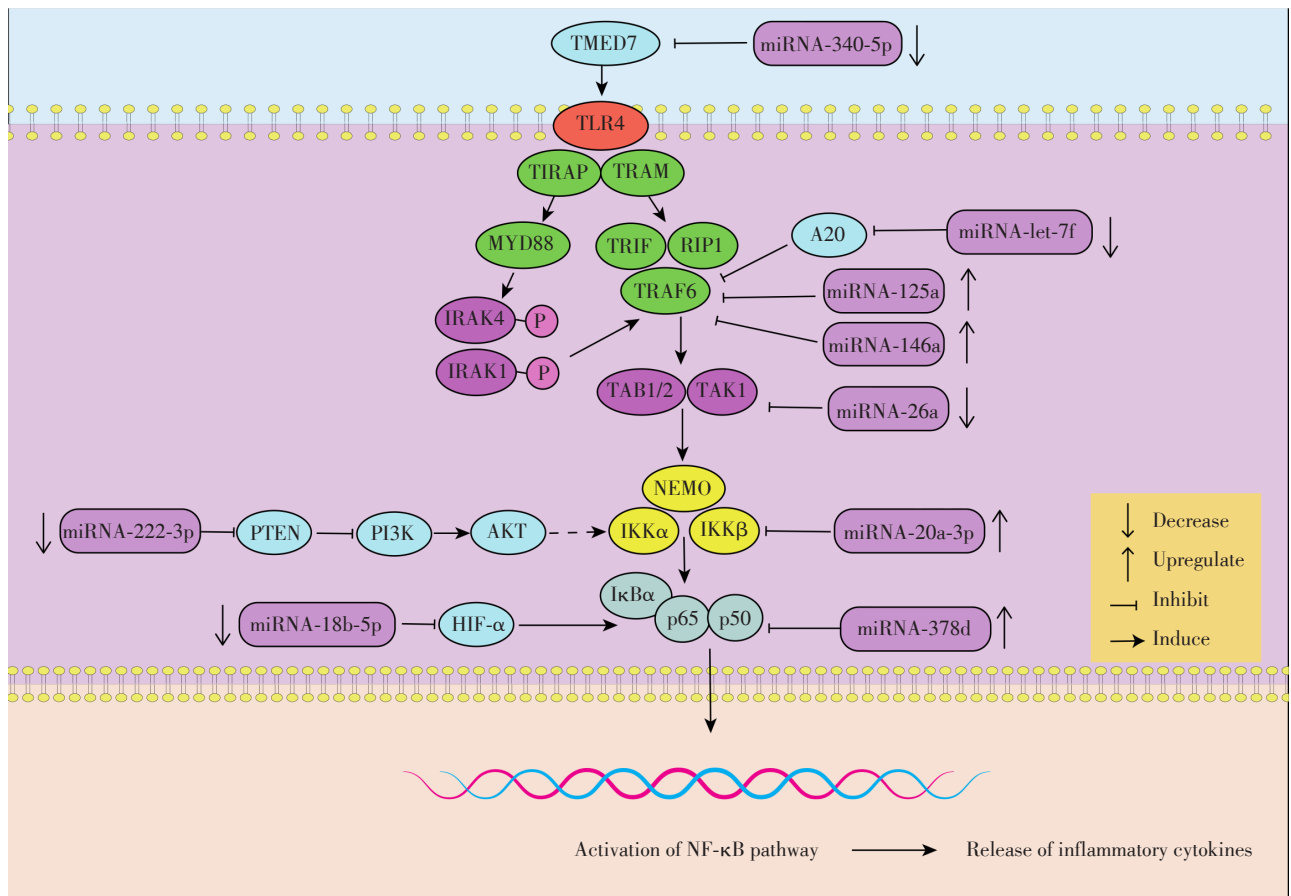


图3 miRNA在感染Mtb后的差异表达及对TLR/NF- κ B通路的影响

Figure 3 The differential expression of miRNAs following Mtb infection and their effect on the TLR/NF- κ B pathway

(transmembrane p24 trafficking protein 7, TMED7)/NF- κ B表达,引起促炎细胞因子的释放^[51]。转化生长因子 β 激活激酶(transforming growth factor β -activated kinase-1, TAK1)是NF- κ B通路中的关键酶。miRNA-26a在感染的细胞中下调,进而导致其靶基因TAK1上调,从而激活NF- κ B信号通路^[52]。在Mtb感染的巨噬细胞中,miRNA-18b-5p表达下调,其靶基因HIF-1 α 表达增加,MAPK p38和NF- κ B p65磷酸化被激活。协同作用导致胞内促炎细胞因子表达上调^[53]。最近一项研究证实,Mtb通过NF- κ B通路导致miRNA-378d的表达下调,且其下调进一步强化了NF- κ B通路的激活,与miRNA-378d靶基因Rab10的上调一起促进促炎细胞因子分泌,抑制胞内Mtb的存活^[54]。

TLR可通过MyD88介导信号转导,MyD88诱导IL-1受体相关激酶(IL-1 receptor associated kinase, IRAK)1和IRAK4的募集,进一步募集TRAF6,激活TAK复合体,导致经典的NF- κ B通路活化^[43]。如前所述,NF- κ B通路直接影响促炎细胞因子的产生,部分miRNA可通过直接靶向TLR影响细胞内的炎症反应。miRNA-21-5p的过表达靶向抑制Bcl-2和TLR4,从而抑制促炎细胞因子的产生^[55]。在Mtb感染巨噬细胞后,miRNA-708-5p水平上调,其通过靶向抑制TLR4降低相关细胞因子的分泌^[56]。miRNA-1178在感染的巨噬细胞中上调,靶向TLR4显著降低了Mtb感染的巨噬细胞中促炎细胞因子的积累^[57]。除了上述通路外,miRNA-196b-5p/SOCS3^[58]、miRNA-370-3p/卵泡抑素样蛋白1(follistatin like 1, FSTL1)^[59]和miRNA-32-5p/FSTL1^[60]以及miRNA-206/金属蛋白酶组织抑制因子3(tissue inhibitor of metalloproteinase 3, TIMP3)^[61]信号均在Mtb感染的细胞中发挥抑制炎症因子的作用。

1.3.2 miRNA调控活性氧(reactive oxygen species, ROS)水平

人巨噬细胞通过增加过氧化物酶体产生ROS来应对感染,限制胞内的Mtb^[62]。Mtb下调巨噬细胞中的miRNA-495,进而上调超氧化物歧化酶2(superoxide dismutase 2, SOD2)的表达,降低ROS水平^[63]。miRNA-346-3p在感染的巨噬细胞中下调,靶向RIPK1/RIPK3/MLKL通路,导致ROS释放和Ca²⁺内流,从而促进Mtb诱导的巨噬细胞坏死^[64]。此外,高菌量活动期TB患者外周血单个核细胞中的miRNA-23a-3p表达下调,通过靶向IRF1/SP1促进ROS生成^[65]。

总之,miRNA通过调节炎症反应和激活细胞因子,在控制Mtb感染中发挥重要作用。了解其间相互作用的机制可以更好地理解Mtb感染的复杂免疫反应,并进一步探究潜在的治疗方法。

2 miRNA作为潜在的治疗靶点

miRNA表达的生理变化对调节复杂的基因网络和细胞信号至关重要。在TB中,miRNA发生病理性的差异表达,导致下游靶基因信号通路失调,影响患者的免疫功能。因而,逆转病理性表达miRNA可增强患者免疫功能,促进Mtb的清除或减轻炎症损伤。miRNA治疗的总体目标是逆转病理性的miRNA表达变化,即人为上调或增强被病理性抑制的内源性miRNA和下调或阻断驱动疾病进展的内源性miRNA。常用于改变miRNA含量的制剂包括:miRNA模拟剂(miRNA mimic)、携带miRNA编码序列的重组表达载体、miRNA抑制剂(anti-miRNA)和miRNA海绵等。鉴于miRNA治疗剂具有较低的细胞膜通透性,可结合纳米载体、病毒转染、引入化学修饰或与生物分子,进一步通过受体介导的摄取来实现胞内递送^[66]。

如前所述,Mtb感染后miRNA含量的改变影响下游分子,进而影响机体抗TB免疫。体外实验中,人为改变相关miRNA的表达可促进抗TB免疫,降低胞内Mtb负荷(表1)。其中,miRNA-155的作用具有争议,报道其上调会促进Mtb的胞内清除,也有研究称其上调会加重感染细胞的细菌负荷。这与前文所述的miRNA-155抗TB作用的双面性相符合^[32],miRNA-155在调节宿主对Mtb感染的免疫反应中具有多种作用,其在Mtb感染不同阶段的靶向作用途径亟待进一步探究。

近年来,许多体内实验已证明了miRNA可能成为潜在的TB治疗靶点,相关生物制剂处理的TB小鼠的疾病进展发生显著变化。miRNA-325^{-/-}小鼠在感染Mtb后,相比野生型小鼠,肺、脾脏和肝脏的载菌量均显著下调,而miR-325^{-/-}小鼠在补充miRNA-325模拟剂后,其器官载菌量显著提升,提示下调TB小鼠的miRNA-325可能有助于缓解TB进展^[35]。miRNA-20b可通过靶向NLRP3/Caspase-1/IL-1 β 通路减轻小鼠的肺部炎症反应,使用miRNA-20b模拟剂处理TB小鼠,其肺部炎症相比对照组明显缓解。相似地,给予miRNA-31激动剂的TB小鼠具有更轻的肺部炎症损伤^[67]。而使用miRNA-25抑制剂处理BCG感染的小鼠,相比对照组,有着更低的肺部

细菌负荷和炎症水平^[18],提示下调 miRNA-25 可以控制 TB 进展。曾有研究者发现用 miRNA-27a 拮抗剂处理的 TB 小鼠,相比对照组有着更低的细菌负荷和肺部炎症水平^[21],以此证明 miRNA-27a 可以作为 TB 治疗的靶点。miRNA-337-3p 靶向 TLR4/MYD88 和 STAT3,导致维生 D 受体介导的抗菌反应受损。miRNA-337-3p 抑制剂处理的 TB 小鼠内脏细菌负荷

显著下调^[68]。此外,miRNA 相关制剂除了可以改善 TB 动物模型的肺部炎症和细菌负荷外,还可改善结核性肺纤维化。miRNA-148a 抑制 NAPDH 氧化酶 4 和 DNA 聚合酶相互作用蛋白 2 表达,miRNA-148a 模拟剂处理 BCG 感染的小鼠后,小鼠胸膜间皮细胞胶原蛋白 1A 的合成受到抑制,从而延缓结核性胸膜炎的纤维化,证明 miRNA-148a 可能有助于缓解结核性

表 1 体外实验证实可降低胞内 Mtb 负荷的 miRNA

Table 1 The miRNAs that reduce the intracellular Mtb load confirmed by *in vitro* experiments

Overexpression of miRNAs that reduce intracellular mycobacterial burden	Downregulation of miRNAs that reduce intracellular mycobacterial burden
miRNA-30a-5p ^[70] , miRNA-20b ^[71] , miRNA-27b ^[39] , miRNA-495 ^[63] , mmu-miRNA-25-3p ^[24] , miRNA-240-5p ^[51] , miRNA-1249-3p ^[72]	miRNA-20b-5p ^[36] , miRNA-106a ^[10] , miRNA-579-3p ^[73] , miRNA-30a-3p ^[70] , miRNA-1301-3p, miRNA-5194 ^[74] , miRNA-18b ^[53] , miRNA-20a-3p ^[45] , miRNA-142-3p ^[9] , miRNA-502-3p ^[50] , miRNA-21-5p ^[55] , miRNA-542-3p ^[16] , miRNA-100-5p ^[40] , miRNA-20a-5p ^[41] , miRNA-27a ^[21] , miRNA-144-3p ^[12] , miRNA-708-5p ^[56] , miRNA-889 ^[20] , miRNA-18a ^[26] , miRNA-25 ^[18] , miRNA-4687-5p ^[75] , miRNA-144* ^[22] , miRNA-1958 ^[15] , miRNA-125b-5p ^[42] , miRNA-370-3p ^[76] , miRNA-378d ^[54] , miRNA-146a ^[47] , miRNA-325-3p ^[35] , miRNA-1178 ^[57] , miRNA-337-3p ^[68] , miRNA-340-5p ^[51] , miRNA-32-5p ^[60] , miRNA-125a ^[48]

肺纤维化^[69]。

因此,miRNA 可以用于增强宿主对 Mtb 的免疫能力,为 TB 的治疗提供了一种潜在方法。许多基于 miRNA 的疗法已经进入临床试验。然而,一种 miRNA 可靶向多个靶基因,一条基因或一条通路通常受到多个 miRNA 的调控,从而形成一个复杂的调控网络。改变一种 miRNA 含量可能会影响多条通路。因而未来还需要进一步的研究来明确 miRNA 调控网络、miRNA 相关药物不良反应、如何使药物有效靶向靶细胞和选择靶向递送药物的材料,从而为基于 miRNA 的抗 TB 靶向药物研发提供理论基础。

3 总结与展望

文章总结了 miRNA 相关抗 TB 免疫调控及其作为潜在治疗靶点的研究进展;综述了 Mtb 感染时,miRNA 含量改变对下游因子的调控以及对自噬、凋亡和炎症反应等生物学过程的影响;进一步提出 miRNA 作为免疫调控因子具有作为 TB 治疗靶点的潜力。

诚然,越来越多的研究揭示了 miRNA 调控宿主免疫反应的作用机制,其中大部分涉及巨噬细胞和先天性免疫应答,但在关于 miRNA 在特定免疫细胞群中的表达谱和调控作用知之甚少,缺乏 miRNA 对适应性免疫应答调控的相关机制。需要进一步明确 miRNA 在感染性疾病中的作用机制。且 miRNA

在体内的表达具有周期性,进一步研究 miRNA 表达谱与 TB 进展的关系有助于更好地理解 TB 的发病机制。此外,目前研究主要集中在单个 miRNA 对细胞分子层面的影响,多个具有不同靶标的 miRNA 如何影响感染过程中的整体宿主反应还有待研究。并且 miRNA 作为 TB 治疗靶点也存在局限。miRNA 用作治疗药物尚处于起步阶段,治疗性 miRNA 如何正确靶向靶细胞是需要攻克的一个难题。鉴于人工智能 (artificial intelligence, AI) 技术的发展,可利用 AI 技术设计合理的寡核苷酸载体和转移载体以减少对非靶细胞的影响。值得注意的是,miRNA 递送材料的选择和设计也会影响 miRNA 的功效,今后还需要更多基于 miRNA 及其相关药物递送系统选择合理性的研究。

总之,miRNA 作为免疫调控分子在调节 Mtb 复制和宿主机体稳态中发挥重要作用,是 TB 的潜在治疗靶点。明确 miRNA 在 Mtb 感染过程中对宿主免疫应答的调控机制,有助于更好地理解 TB 的发病机制及开发相应的靶向治疗药物。

[参考文献]

- [1] WORLD HEALTH ORGANIZATION. Global tuberculosis report 2023[R]. Geneva: World Health Organization, 2023
- [2] 陈 玲,胡春梅,余 敏,等. 肺癌合并肺结核的机制研究及诊治进展[J]. 南京医科大学学报(自然科学版),

- 2022, 42(5): 746–750, 758
- [3] ALIPOOR S D, ADCOCK I M, TABARSI P, et al. MiRNAs in tuberculosis: their decisive role in the fate of TB[J]. *Eur J Pharmacol*, 2020, 886: 173529
- [4] IWAKAWA H O, TOMARI Y. Life of RISC: formation, action, and degradation of RNA-induced silencing complex [J]. *Mol Cell*, 2022, 82(1): 30–43
- [5] DERETIC V. Autophagy in inflammation, infection, and immunometabolism[J]. *Immunity*, 2021, 54(3): 437–453
- [6] GOLOVKINE G R, ROBERTS A W, MORRISON H M, et al. Autophagy restricts *Mycobacterium tuberculosis* during acute infection in mice [J]. *Nat Microbiol*, 2023, 8(5): 819–832
- [7] SHARIQ M, QUADIR N, ALAM A, et al. The exploitation of host autophagy and ubiquitin machinery by *Mycobacterium tuberculosis* in shaping immune responses and host defense during infection [J]. *Autophagy*, 2023, 19(1): 3–23
- [8] LEVINE B, KROEMER G. Biological functions of autophagy genes: a disease perspective [J]. *Cell*, 2019, 176(1/2): 11–42
- [9] QU Y L, GAO Q, WU S, et al. MicroRNA-142-3p inhibits autophagy and promotes intracellular survival of *Mycobacterium tuberculosis* by targeting ATG16L1 and ATG4c [J]. *Int Immunopharmacol*, 2021, 101(Pt A): 108202
- [10] LIU K M, HONG D T, ZHANG F, et al. MicroRNA-106a inhibits autophagy process and antimicrobial responses by targeting ULK1, ATG7, and ATG16L1 during mycobacterial infection [J]. *Front Immunol*, 2020, 11: 610021
- [11] GUO L, ZHAO J, QU Y L, et al. MicroRNA-20a inhibits autophagic process by targeting ATG7 and ATG16L1 and favors mycobacterial survival in macrophage cells [J]. *Front Cell Infect Microbiol*, 2016, 6: 134
- [12] GUO L, ZHOU L L, GAO Q, et al. MicroRNA-144-3p inhibits autophagy activation and enhances *Bacillus Calmette-Guérin* infection by targeting ATG4a in RAW264.7 macrophage cells [J]. *PLoS One*, 2017, 12(6): e0179772
- [13] WU J, ZHANG Y, TANG H, et al. MicroRNA-144-3p inhibits host lipid catabolism and autophagy by targeting PPAR α and ABCA1 during *Mycobacterium tuberculosis* infection [J]. *ACS Infect Dis*, 2024, 10(5): 1654–1663
- [14] ETNA M P, SINIGAGLIA A, GRASSI A, et al. *Mycobacterium tuberculosis*-induced miR-155 subverts autophagy by targeting ATG3 in human dendritic cells [J]. *PLoS Pathog*, 2018, 14(1): e1006790
- [15] DING S Q, QU Y L, YANG S Q, et al. Novel miR-1958 promotes *Mycobacterium tuberculosis* survival in RAW264.7 cells by inhibiting autophagy via Atg5 [J]. *J Microbiol Biotechnol*, 2019, 29(6): 989–998
- [16] LUO D, WU J L, LIU Y Y, et al. Overexpression of VPS11 antagonizes the promoting effect of miR-542-3p on *Mycobacterium tuberculosis* survival in macrophages by regulating autophagy [J]. *Microb Pathog*, 2022, 169: 105609
- [17] TU H H, YANG S, JIANG T T, et al. Elevated pulmonary tuberculosis biomarker miR-423-5p plays critical role in the occurrence of active TB by inhibiting autophagosome-lysosome fusion [J]. *Emerg Microbes Infect*, 2019, 8(1): 448–460
- [18] DONG W Q, WANG G Y, FENG J J, et al. MiR-25 blunts autophagy and promotes the survival of *Mycobacterium tuberculosis* by regulating NPC1 [J]. *iScience*, 2022, 25(5): 104279
- [19] BEHURA A, MISHRA A, CHUGH S, et al. ESAT-6 modulates calcimycin-induced autophagy through microRNA-30a in mycobacteria infected macrophages [J]. *J Infect*, 2019, 79(2): 139–152
- [20] CHEN D Y, CHEN Y M, LIN C F, et al. MicroRNA-889 inhibits autophagy to maintain mycobacterial survival in patients with latent tuberculosis infection by targeting TWEAK [J]. *mBio*, 2020, 11(1): e03045–e03049
- [21] LIU F, CHEN J X, WANG P, et al. MicroRNA-27a controls the intracellular survival of *Mycobacterium tuberculosis* by regulating calcium-associated autophagy [J]. *Nat Commun*, 2018, 9(1): 4295
- [22] KIM J K, LEE H M, PARK K S, et al. MIR144* inhibits antimicrobial responses against *Mycobacterium tuberculosis* in human monocytes and macrophages by targeting the autophagy protein DRAM2 [J]. *Autophagy*, 2017, 13(2): 423–441
- [23] DENG F, XU P, MIAO J H, et al. Pulmonary tuberculosis biomarker miR-215-5p inhibits autophagosome-lysosome fusion in macrophages [J]. *Tuberculosis*, 2023, 143: 102422
- [24] YUAN W Q, ZHAN X H, LIU W, et al. Mmu-miR-25-3p promotes macrophage autophagy by targeting DUSP10 to reduce mycobacteria survival [J]. *Front Cell Infect Microbiol*, 2023, 13: 1120570
- [25] WU M, LIU Z B, ZHANG S J. Down-regulation of hsa_circ_0045474 induces macrophage autophagy in tuberculosis via miR-582-5p/TNKS2 axis [J]. *Innate Immun*, 2022, 28(1): 11–18
- [26] YUAN Q L, CHEN H T, YANG Y X, et al. MiR-18a promotes *Mycobacterial* survival in macrophages via inhibiting autophagy by down-regulation of ATM [J]. *J Cell Mol Med*, 2020, 24(2): 2004–2012
- [27] SONG J, SUN J G, WANG Y Q, et al. CeRNA network identified hsa-miR-17-5p, hsa-miR-106a-5p and hsa-miR-2355-5p as potential diagnostic biomarkers for tuberculo-

- sis[J]. *Medicine*, 2023, 102(11): e33117
- [28] ARNETT E, PAHARI S, LEOPOLD W C M, et al. Combination of MCL-1 and BCL-2 inhibitors is a promising approach for a host-directed therapy for tuberculosis [J]. *Biomedicine Pharmacother*, 2023, 168: 115738
- [29] STUTZ M D, ALLISON C C, OJAIMI S, et al. Macrophage and neutrophil death programs differentially confer resistance to tuberculosis [J]. *Immunity*, 2021, 54(8): 1758-1771
- [30] NISA A, KIPPER F C, PANIGRAHY D, et al. Different modalities of host cell death and their impact on *Mycobacterium tuberculosis* infection [J]. *Am J Physiol Cell Physiol*, 2022, 323(5): C1444-C1474
- [31] BONILLA-MURO M G, HERNÁNDEZ DE LA CRUZ O N, GONZALEZ-BARRIOS J A, et al. EsxA mainly contributes to the miR-155 overexpression in human monocyte-derived macrophages and potentially affect the immune mechanism of macrophages through miRNA dysregulation [J]. *J Microbiol Immunol*, 2021, 54(2): 185-192
- [32] ABDALLA A E, ALANAZI A, ABOSALIF K O A, et al. MicroRNA-155, a double-blade sword regulator of innate tuberculosis immunity [J]. *Microb Pathog*, 2023, 185: 106438
- [33] HUANG J, JIAO J H, XU W H, et al. MiR-155 is upregulated in patients with active tuberculosis and inhibits apoptosis of monocytes by targeting FOXO3 [J]. *Mol Med Rep*, 2015, 12(5): 7102-7108
- [34] YANG S, LI F, JIA S, et al. Early secreted antigen ESAT-6 of *Mycobacterium tuberculosis* promotes apoptosis of macrophages via targeting the microRNA155-SOCS1 interaction [J]. *Cell Physiol Biochem*, 2015, 35(4): 1276-1288
- [35] FU B B, XUE W W, ZHANG H W, et al. MicroRNA-325-3p facilitates immune escape of *Mycobacterium tuberculosis* through targeting LNX1 via NEK6 accumulation to promote anti-apoptotic STAT3 signaling [J]. *mBio*, 2020, 11(3): e00557-20
- [36] ZHANG D F, YI Z J, FU Y R. Downregulation of miR-20b-5p facilitates *Mycobacterium tuberculosis* survival in RAW 264.7 macrophages via attenuating the cell apoptosis by Mcl-1 upregulation [J]. *J Cell Biochem*, 2019, 120(4): 5889-5896
- [37] YUAN S, WU Q, WANG Z W, et al. MiR-223: an immune regulator in infectious disorders [J]. *Front Immunol*, 2021, 12: 781815
- [38] FU B B, LIN X Y, TAN S, et al. MiR-342 controls *Mycobacterium tuberculosis* susceptibility by modulating inflammation and cell death [J]. *EMBO Rep*, 2021, 22(9): e52252
- [39] LIANG S X, SONG Z G, WU Y Y, et al. MicroRNA-27b modulates inflammatory response and apoptosis during *Mycobacterium tuberculosis* infection [J]. *J Immunol*, 2018, 200(10): 3506-3518
- [40] SU L, ZHU T T, LIU H, et al. The miR-100-5p targets SMARCA5 to regulate the apoptosis and intracellular survival of BCG in infected THP-1 cells [J]. *Cells*, 2023, 12(3): 476
- [41] ZHANG G L, LIU X, WANG W F, et al. Down-regulation of miR-20a-5p triggers cell apoptosis to facilitate mycobacterial clearance through targeting JNK2 in human macrophages [J]. *Cell Cycle*, 2016, 15(18): 2527-2538
- [42] LIU G M, WAN Q F, LI J W, et al. Silencing miR-125b-5p attenuates inflammatory response and apoptosis inhibition in *Mycobacterium tuberculosis*-infected human macrophages by targeting DNA damage-regulated autophagy modulator 2 (DRAM2) [J]. *Cell Cycle*, 2020, 19(22): 3182-3194
- [43] TIWARI D, MARTINEAU A R. Inflammation-mediated tissue damage in pulmonary tuberculosis and host-directed therapeutic strategies [J]. *Semin Immunol*, 2023, 65: 101672
- [44] ZHOU X Y, ZHANG L J, LIE L M, et al. MxA suppresses TAK1-IKK α/β -NF- κ B mediated inflammatory cytokine production to facilitate *Mycobacterium tuberculosis* infection [J]. *J Infect*, 2020, 81(2): 231-241
- [45] CUI J W, LI Z Y, CUI K L, et al. MicroRNA-20a-3p regulates the host immune response to facilitate the *Mycobacterium tuberculosis* infection by targeting IKK β /NF- κ B pathway [J]. *Int Immunopharmacol*, 2021, 91: 107286
- [46] KUMAR M, SAHU S K, KUMAR R, et al. MicroRNA let-7 modulates the immune response to *Mycobacterium tuberculosis* infection via control of A20, an inhibitor of the NF- κ B pathway [J]. *Cell Host Microbe*, 2015, 17(3): 345-356
- [47] LI M, WANG J L, FANG Y M, et al. MicroRNA-146a promotes mycobacterial survival in macrophages through suppressing nitric oxide production [J]. *Sci Rep*, 2016, 6: 23351
- [48] NIU W Y, SUN B, LI M Y, et al. TLR-4/microRNA-125a/NF- κ B signaling modulates the immune response to *Mycobacterium tuberculosis* infection [J]. *Cell Cycle*, 2018, 17(15): 1931-1945
- [49] CHEN Z H, LUO T, MA P J, et al. *Mycobacterium tuberculosis* ESAT6 modulates host innate immunity by downregulating miR-222-3p target PTEN [J]. *Biochim Biophys Acta Mol Basis Dis*, 2022, 1868(1): 166292
- [50] LIU F, DONG Z, LIN Y F, et al. MicroRNA-502-3p promotes *Mycobacterium tuberculosis* survival in macrophages by modulating the inflammatory response by tar-

- geting ROCK1[J]. Mol Med Rep, 2021, 24(5): 753
- [51] ZHANG B L, LI H L, ZHANG J L, et al. Overexpression of microRNA-340-5p ameliorates inflammatory response and intracellular survival of *Mycobacterium tuberculosis* in alveolar type II cells[J]. Infect Drug Resist, 2021, 14: 1573–1584
- [52] LI H W, WANG Y F, SONG Y Z. MicroRNA-26b inhibits the immune response to *Mycobacterium tuberculosis* (M.tb) infection in THP-1 cells via targeting TGF β -activated kinase-1 (TAK1), a promoter of the NF- κ B pathway[J]. Int J Clin Exp Pathol, 2018, 11(3): 1218–1227
- [53] ZHU T, LIU H, SU L, et al. MicroRNA-186-5p downregulation favors *Mycobacterium tuberculosis* clearance in macrophages via HIF-1 α by promoting an inflammatory response[J]. ACS Infect Dis, 2021, 7(4): 800–810
- [54] ZHU Y F, XIAO Y, KONG D L, et al. Down-regulation of miR-378d increased Rab10 expression to help clearance of *Mycobacterium tuberculosis* in macrophages[J]. Front Cell Infect Microbiol, 2020, 10: 108
- [55] ZHAO Z H, HAO J Z, LI X, et al. MiR-21-5p regulates mycobacterial survival and inflammatory responses by targeting Bel-2 and TLR4 in *Mycobacterium tuberculosis*-infected macrophages[J]. FEBS Lett, 2019, 593(12): 1326–1335
- [56] LI W T, ZHANG Q. MicroRNA-708-5p regulates mycobacterial vitality and the secretion of inflammatory factors in *Mycobacterium tuberculosis*-infected macrophages by targeting TLR4[J]. Eur Rev Med Pharmacol Sci, 2019, 23(18): 8028–8038
- [57] SHI G, MAO G F, XIE K J, et al. MiR-1178 regulates mycobacterial survival and inflammatory responses in *Mycobacterium tuberculosis*-infected macrophages partly via TLR4[J]. J Cell Biochem, 2018, 119(9): 7449–7457
- [58] YUAN Y Q, LIN D Z, FENG L, et al. Upregulation of miR-196b-5p attenuates BCG uptake via targeting SOCS3 and activating STAT3 in macrophages from patients with long-term cigarette smoking-related active pulmonary tuberculosis[J]. J Transl Med, 2018, 16(1): 284
- [59] ZHANG Y, LUO D L, TANG M L, et al. Circ-WDR27 regulates mycobacterial vitality and secretion of inflammatory cytokines in *Mycobacterium tuberculosis*-infected macrophages via the miR-370-3p/FSTL1 signal network[J]. J Biosci, 2022, 47: 28
- [60] ZHANG Z M, ZHANG A R, XU M, et al. TLR-4/miRNA-32-5p/FSTL1 signaling regulates mycobacterial survival and inflammatory responses in *Mycobacterium tuberculosis*-infected macrophages[J]. Exp Cell Res, 2017, 352(2): 313–321
- [61] FU X D, ZENG L H, LIU Z, et al. MicroRNA-206 regulates the secretion of inflammatory cytokines and MMP9 expression by targeting TIMP3 in *Mycobacterium tuberculosis*-infected THP-1 human macrophages[J]. Biochem Biophys Res Commun, 2016, 477(2): 167–173
- [62] PELLEGRINO E, AYLAN B, BUSSI C, et al. Peroxisomal ROS control cytosolic *Mycobacterium tuberculosis* replication in human macrophages[J]. J Cell Biol, 2023, 222(12): e202303066
- [63] REN X X, DONG W Q, FENG J J, et al. MiR-495 regulates cellular reactive oxygen species levels by targeting sod2 to inhibit intracellular survival of *Mycobacterium tuberculosis* in macrophages[J]. Infect Immun, 2021, 89(12): e0031521
- [64] LIU L, YU Z R, MA Q M, et al. LncRNA NR_003508 suppresses *Mycobacterium tuberculosis*-induced programmed necrosis via sponging miR-346-3p to regulate RIPK1[J]. Int J Mol Sci, 2023, 24(9): 8016
- [65] CHEN Y C, LEE C P, HSIAO C C, et al. MicroRNA-23a-3p down-regulation in active pulmonary tuberculosis patients with high bacterial burden inhibits mononuclear cell function and phagocytosis through TLR4/TNF- α /TGF- β 1/IL-10 signaling via targeting IRF1/SP1[J]. Int J Mol Sci, 2020, 21(22): 8587
- [66] DIENER C, KELLER A, MEESE E. Emerging concepts of miRNA therapeutics: from cells to clinic[J]. Trends Genet, 2022, 38(6): 613–626
- [67] ZHANG Z Y, MAI Q D, YANG L J, et al. MicroRNA-31 mediated by interferon regulatory factor 7 signaling facilitates control of *Mycobacterium tuberculosis* infection[J]. Int J Med Microbiol, 2022, 312(7): 151569
- [68] LIANG S S, HUANG G X, WU T, et al. MIR337-3p enhances mycobacterial pathogenicity involving TLR4/MYD88 and STAT3 signals, impairing VDR antimicrobial response and fast-acting immunity[J]. Front Immunol, 2021, 12: 739219
- [69] WOO S J, KIM Y, JUNG H, et al. MicroRNA 148a suppresses tuberculous fibrosis by targeting NOX4 and POL-DIP2[J]. Int J Mol Sci, 2022, 23(6): 2999
- [70] BEHURA A, DAS M, KUMAR A, et al. ESAT-6 impedes IL-18 mediated phagosome lysosome fusion via microRNA-30a upon calcimycin treatment in mycobacteria infected macrophages[J]. Int Immunopharmacol, 2021, 101 (Pt A): 108319
- [71] YAN K, XU G, LI Z. MicroRNA-20b carried by mesenchymal stem cell-derived extracellular vesicles protects alveolar epithelial type II cells from *Mycobacterium tuberculosis* infection *in vitro* [J]. Infect Genet Evol, 2022, 101: 105292

- [61] MARYAM, VARGHESE T P, TAZNEEM B. Unraveling the complex pathophysiology of heart failure: insights into the role of renin-angiotensin-aldosterone system (RAAS) and sympathetic nervous system (SNS) [J]. *Curr Probl Cardiol*, 2024, 49(4): 102411
- [62] VAN DER BIJL P, KNUUTI J, DELGADO V, et al. Cardiac Sympathetic Innervation Imaging with PET Radiotracers [J]. *Curr Cardiol Rep*, 2020, 23(1): 4
- [63] 张敏, 何玉林, 王相成, 等. ¹¹C标记的心脏交感神经受体显像剂的研究进展 [J]. *国际放射医学核医学杂志*, 2023, 47(2): 105-111
- [64] FALLAVOLLITA J A, HEAVEY B M, LUISI A J, et al. Regional myocardial sympathetic denervation predicts the risk of sudden cardiac arrest in ischemic cardiomyopathy [J]. *J Am Coll Cardiol*, 2014, 63(2): 141-149
- [65] AIKAWA T, NAYA M, OBARA M, et al. Impaired myocardial sympathetic innervation is associated with diastolic dysfunction in heart failure with preserved ejection fraction: ¹¹C-hydroxyephedrine PET study [J]. *J Nucl Med*, 2017, 58(5): 784-790
- [66] GUPTA S, GE Y, SINGH A, et al. Multimodality imaging assessment of myocardial fibrosis [J]. *JACC Cardiovasc Imaging*, 2021, 14(12): 2457-2469
- [67] VARASTEHEH Z, MOHANTA S, ROBU S, et al. Molecular imaging of fibroblast activity after myocardial infarction using a ⁶⁸Ga-labeled fibroblast activation protein inhibitor, FAPI-04 [J]. *J Nucl Med*, 2019, 60(12): 1743-1749
- [68] DIEKMANN J, KOENIG T, ZWADLO C, et al. Molecular imaging identifies fibroblast activation beyond the infarct region after acute myocardial infarction [J]. *J Am Coll Cardiol*, 2021, 77(14): 1835-1837
- [69] HIGUCHI T, SERFLING S E, LEISTNER D M, et al. FAPI-PET in cardiovascular disease [J]. *Semin Nucl Med*, 2024, 54(5): 747-752

[收稿日期] 2024-07-19

(本文编辑: 唐震)

(上接第1754页)

- [72] MA F Q, WANG X, QIU Z H, et al. NK-derived exosome miR-1249-3p inhibits *Mycobacterium tuberculosis* survival in macrophages by targeting SKOR1 [J]. *Cytokine*, 2024, 175: 156481
- [73] DENG Q, HUANG J, YAN J J, et al. Circ_0001490/miR-579-3p/FSTL1 axis modulates the survival of mycobacteria and the viability, apoptosis and inflammatory response in *Mycobacterium tuberculosis*-infected macrophages [J]. *Tuberculosis*, 2021, 131: 102123
- [74] QU Y L, JIANG D, LIU M J, et al. LncRNA DANCR restrained the survival of *Mycobacterium tuberculosis* H37Ra by sponging miR-1301-3p/miR-5194 [J]. *Front Microbiol*, 2023, 14: 1119629
- [75] MENG C Q, CHEN G X, LIU Y, et al. MiR-4687-5p affects intracellular survival of *Mycobacterium tuberculosis* through its regulation of NRAMP1 expression in A549 cells [J]. *Microorganisms*, 2024, 12(1): 227
- [76] TAMGUE O, GCANGA L, OZTURK M, et al. Differential targeting of c-maf, bach-1, and elmo-1 by microRNA-143 and microRNA-365 promotes the intracellular growth of *Mycobacterium tuberculosis* in alternatively IL-4/IL-13 activated macrophages [J]. *Front Immunol*, 2019, 10: 421

[收稿日期] 2024-06-26

(本文编辑: 陈汐敏)